# Relative bioavailability of rafoxanide following intraruminal and intra-abomasal administration in sheep

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# **ABSTRACT**

The bioavailability of rafoxanide was compared after intraruminal and intra-abomasal administration in healthy adult sheep (n=6) in a single dose, 2 parallel group study at 7.5 mg/kg. Rafoxanide concentrations in plasma were measured by means of HPLC analysis. Primary pharmacokinetic parameters for bioavailability and disposition of rafoxanide in plasma for both routes of administration were determined by noncompartmental and non-linear, 1-compartmental pharmacokinetic analysis, respectively. Significantly ( $P \le 0.05$ ) higher peak plasma concentrations ( $c_{max}$ ) of rafoxanide and a more rapid rate of absorption (c. 3.5 times) was observed in sheep after intra-abomasal (i-a) administration compared to intraruminal (i.r.) administration. A significantly ( $P \le 0.05$ ) longer lag period ( $t_{lag}$ ) before absorption ( $6.8 \pm 2.9$  h) occurred after i.r. than after i-a treatment ( $1.9 \pm 0.6$  h). There was no significant difference (P > 0.05) in AUC, MRT and in the rates of elimination ( $t_{10}$ -HL and  $t_{1/2}$ ) between the i.r. and i-a routes of administration. The results of the study demonstrated the important influence of the rumino-reticulum on absorption of rafoxanide in sheep.

**Key words**: intra-abomasal, intraruminal, pharmacokinetics, rafoxanide, salicylanilides, sheep.

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# INTRODUCTION

Rafoxanide (Ranide, Logos Agvet) belongs to the group of halogenated salicylanilide anthelmintic agents used extensively for the control of liver fluke and blood-sucking nematodes in sheep and cattle, and larvae of Oestrus ovis in sheep. Halogenated salicylanilides, in particular rafoxanide and closantel, share similar chemical, pharmacokinetic (extensive plasma protein binding and long elimination half-life) and safety features<sup>30</sup>. The efficacy of rafoxanide and closantel against bloodsucking parasites and the persistent anthelmintic effect of closantel have been directly associated with the pharmacokinetics of these agents<sup>3,12,13,18,21,22,24</sup>. Rafoxanide concentrations in plasma have also been positively correlated with the occurrence of toxicity in lambs<sup>33</sup>, which in addition substantiates the important relationship between the pharmacokinetics of these drugs and their safety. Apart from a few studies 9,15,22,31-33, there has been little pharmacokinetic focus on the absorption and disposition of the halogenated salicylanilides. The intravascular disposition of closantel and rafoxanide has only recently been reported 30.

The absorption of drugs administered orally to ruminants is markedly affected by the anatomical and physiological features of the forestomachs 5,10,16. Absorption from the rumino-reticulum has been reported for several drugs 14,17,22,26, since the ruminal epithelium does not constitute a barrier for the distribution of lipidsoluble, non-polar, hydrophilic substances<sup>10</sup>. Absorption of drugs from the rumen, on the other hand, is generally negligible, owing to the slow rate of diffusion across the ruminal epithelium relative to the rate of outflow from the rumino-reticulum to the abomasum<sup>5</sup>. Poor mixing of drugs in the aqueous phase of rumen digesta, relatively low surface area to volume ratio, proportionally lower blood supply to the rumen epithelium and adsorption of drug particles to particulate matter in the ruminoreticulum are the main causes of the slow rate of diffusion<sup>5,20</sup>. The dilution effect of the large volume of ruminal contents

reduces the diffusion gradient between rumen and plasma. Many drugs are rapidly and extensively adsorbed onto the solid phase of rumen digesta, thereby delaying absorption from the rumen and increasing the proportion of drug outflow to the abomasum and lower gastrointestinal tract<sup>5,14,16</sup>.

Foreign compounds may be inactivated or activated by reduction, hydrolytic and fission metabolic reactions by microflora or by the reducing conditions within the rumino-reticulum fluid <sup>10,28</sup>. They are therefore important considerations in the pharmacokinetics and pharmacodynamics as well as the toxicity of drugs administered orally to ruminants. The halogenated salicylanilide clioxanide is deacetylated in the rumen to form an active hydroxyl derivative<sup>24</sup>. Its anthelmintic activity in sheep is reduced if it is passed directly into the abomasum<sup>6,34</sup>.

Very few studies, other than those on clioxanide, have been conducted to establish the importance of the ruminoreticulum on the absorption of halogenated salicylanilides. Taylor et al. 35 showed that significantly lower peak plasma concentrations and extent of absorption of rafoxanide administered orally occurred in grazing lambs, owing to reduced digesta flow rate, compared to housed lambs that were fed hay. A significantly greater bioavailability of rafoxanide (2.5-3.0 times) administered orally was reported in suckling lambs (5-8 weeks of age) compared to weaned lambs (5 months of age)<sup>32</sup>. The increased absorption was ascribed to the presence of an underdeveloped rumen in the suckling lambs compared to the weaned lambs and a more rapid and extensive absorption from the abomasum. Ruminants at the age of 3–8 weeks are regarded as being within the transitional phase of forestomach development8. Comparison of the efficacy of rafoxanide at 3.75 mg/kg, administered either intra-abomasally (i-a), orally (p.o.) or intraruminally (i.r.), against 6-week-old Fasciola hepatica, revealed that i-a treatment was more than twice as effective than either p.o. or i.r. treatment<sup>7</sup>. Similar efficacy was shown after p.o. and i.r. administration.

No pharmacokinetic studies have been

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reported with rafoxanide administered either i.r. or i-a to demonstrate the difference in bioavailability between the 2 routes. In the current study bioavailability of rafoxanide was compared after i-a and i.r. routes of administration in adult sheep.

### **MATERIALS AND METHODS**

#### Study design

A randomised balanced, single dose, parallel 2-group, comparative bioavailability study with rafoxanide administered either i-a or i.r., was conducted in 6 healthy adult South African mutton Merino sheep. Equal numbers of ewes (n=3) and wethers (n=3) were used. Whole blood counts, haematocrit, serum aspartate transaminase activity and serum creatinine, urea and albumin concentrations of all animals, determined at the start of the study, were within the normal ranges expected for sheep<sup>4,29</sup>.

An i-a catheter was inserted under general anaesthesia (halothane: Fluothane, ICI) in all sheep 7 days before the start of the trial. A stab incision was made into the abomasum and a feeding catheter inserted through the last intercostal space at the costochondral junction. To prevent leakage of abomasal content, a pursestring was applied at the site of incision. The catheter was stabilised in a channel of abomasal serosa, *c.* 2 cm in length, by means of an external suture.

Rafoxanide 3 % m/v oral suspension (Ranide®, Logos AgVet) at 7.5 mg/kg was administered either i-a and i.r.. Intraabomasal treatment was administered by means of the i-a catheter and i.r. treatment by injection through the left abdominal wall into the rumen.

Venous blood was collected in  $10 \text{ m}\ell$  heparinised vacuum tubes, immediately before treatment and at 0.5, 1, 2, 3, 5, 7, 9, 12, 15, 24 snd 48 h, and 3, 5, 7 10, 14, 21, 28 and 35 days after treatment. Timed collection occurred within 30 sec of the scheduled time up to 24 h, and thereafter within 1-5 min.

Blood samples were centrifuged at 3000 r.p.m. for 15 min and the plasma collected. Two equal aliquots of plasma from each animal were transferred into clean polycarbonate tubes and stored at  $-20\,^{\circ}\mathrm{C}$  until analysed. Recovery and storage of plasma occurred within 12 h of blood collection.

#### Rafoxanide analyses

Rafoxanide plasma concentrations were determined by high-pressure liquid chromatography (HPLC) according to the method described by M Mülders and coworkers (Department of Pharmacology and Toxicology, Faculty of Veterinary Science, University of Pretoria, pers. comm., 1998).

### Pharmacokinetic analysis

Non-compartmental analysis of the plasma concentration versus time data of rafoxanide for extravascular input was performed by PC Nonlin Version 4.2 (Statistical Consultants, New York) computer programme. The area under the plasma concentration versus time curve (AUC, zero-moment) and the 1st non-normalised moment (AUMC) were calculated according to the trapezoidal method from time zero to the last sampling time<sup>11</sup>. Extrapolation of AUC to infinity (AUCinf) was performed using the slope  $(\beta)$  of the terminal phase. The slope  $(\beta)$  of the terminal phase of the curve was determined by linear regression analysis of the terminal plasma concentrations of the semilogarithmic plasma concentration versus time curve. Mean residence time (MRT, 1st moment) was derived from AUC/AUMC. Maximum plasma concentration (c<sub>max</sub>) of either rafoxanide and time to  $c_{max}$  ( $t_{max}$ ) were read directly from the individual plasma concentrations.

Non-linear compartmental analysis of the rafoxanide plasma data was performed using the same pharmacokinetic computer programme using the Nelder-Mead algorithm<sup>23</sup>. Initial pharmacokinetic parameter estimates, used for the non-linear analysis, were derived automatically by initial linear analysis performed by the programme. Akaike's information criterion<sup>36</sup>, based on the mean values of the final estimates of the associated pharmacokinetic parameters and lack of systematic deviations around the fitted disposition curve, was used to determine the number of exponential terms that best described the data. Primary pharmacokinetic parameters were derived by 1-compartmental analysis with 1st-order input, 1st-order output and lag time of the plasma concentration-time data for each individual animal yielding the microconstants  $k_{01}$  and  $k_{10}$ . Secondary disposition parameters, including  $k_{01}$  half-life ( $k_{01}$ -HL) and  $k_{10}$ half-life (k<sub>10</sub>-HL) were derived from the primary parameters utilising standard procedures<sup>11</sup>.

# Statistical analyses

The descriptive statistics (mean  $\pm$  SD) for treatments and treatment groups were calculated for all pharmacokinetic parameters within each study. Differences in the mean pharmacokinetic parameters were statistically compared using the Student's t-test, whereas the nonparametric Wilcoxon rank test was

applied to the rate constants ( $k_{01}$  and  $k_{10}$ ), rate constant half-lives ( $k_{01}$ -HL and  $k_{10}$ -HL),  $t_{max}$  and  $t_{lag}$ . All statistical procedures were performed using the SAS statistical software programme for Windows  $95^1$ .

#### **RESULTS**

The mean plasma concentration *versus* time profiles for rafoxanide following i.r. and i-a administration are illustrated in Fig. 1. The data was best described by a 1-compartmental open model with 1st-order rate constants and lag-time.

Rafoxanide suspension administered i-a reached significantly (P < 0.05) higher peak plasma concentrations compared to i.r. administration in c. 25 % of the time (Table 1). The rate of absorption of rafoxanide after i-a administration was significantly (P < 0.05) more rapid as measured by  $t_{\rm max}$ ,  $t_{\rm lag}$ , and  $k_{01}$ -HL than i.r. treatment. There were no significant differences (P > 0.05) in AUC, MRT and rates of elimination (k10-HL and  $t^{1/2}$ <sub> $\beta$ </sub>) between the 2 routes of administration.

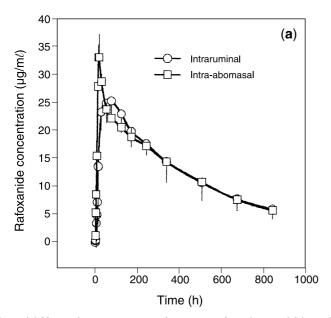
## **DISCUSSION**

The general pharmacokinetic profile of rafoxanide after i.r. and i-a administration observed in this study is consistent with the pharmacokinetics of rafoxanide reported previously in sheep  $^{22,31,32}$ . Very similar AUC,  $c_{max}$ , and  $t\frac{1}{2}$  results were obtained for rafoxanide after i.r. administration compared with those reported by Mohammed-Ali and Bogan following oral treatment in sheep.

The results of the current study indicate the effect of the rumino-reticulum on the absorption of rafoxanide following i.r. administration in adult sheep. Rafoxanide administered i.r. was absorbed significantly slower than after i-a administration and had an extended lag-time before absorption.

According to Bogan and Marriner<sup>5</sup>, the rumino-reticulum serves mainly as a 'reservoir' for drugs and is responsible for comparatively negligible absorption relative to the rest of the gastrointestinal tract. The absorption of drugs administered i.r. is delayed as a result of the dilution effect of the large volume of ruminal contents and adsorption onto rumen digesta resulting in a delay of outflow from the rumino-reticulum<sup>5,14,16</sup>.

Unlike the benzimidazole anthelmintics and ivermectin, the bioavailability of rafoxanide was similar for both i.r and i-a route of administration. The higher  $c_{max}$  plasma concentrations of rafoxanide observed in the sheep after i-a treatment is related to the more rapid rate of absorption from the abomasum and not due to an increase in bioavailability. In the case of



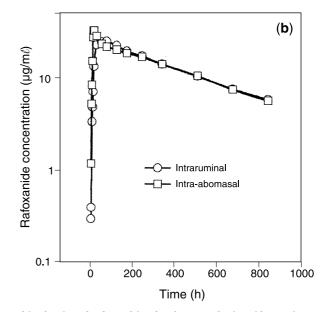


Fig. 1: (a) Mean plasma concentration *versus* time data and (b) semilogarithmic plot of rafoxanide after intraruminal and intra-abomasal administration.

Table 1: Comparative bioavailability and disposition of rafoxanide after intraruminal (i.r.) and intra-abomasal (i.a.) administration in sheep.

Parameter	Mean ± SD	
	i.r. (n = 3)	i.a. (n = 3)
AUClast (μg/h/mℓ)	11414 ± 2080 <sup>a</sup>	11451 ± 80 <sup>a</sup>
AUCinf (μg/h/mℓ)	14892 ± 3541 <sup>a</sup>	14818 ± 313 <sup>a</sup>
$c_{max}$ (µg/m $\ell$ )	$27.5 \pm 2.4^{a}$	$34.6 \pm 3.5^{b}$
k <sub>01</sub> (per h)	$0.1270 \pm 0.0577^{a}$	$0.3976 \pm 0.0560^{b}$
k <sub>10</sub> (per h)	$0.0021 \pm 0.0004^{a}$	$0.0023 \pm 0.0001^a$
k <sub>01</sub> -HL (h)	$6.5 \pm 2.3^{a}$	$1.8 \pm 0.23^{b}$
k <sub>10</sub> -HL (d)	$14.3 \pm 2.5^{a}$	$12.6 \pm 0.8^{a}$
t <sub>max</sub> (h)	$44.0 \pm 24.7^{a}$	11.0 ± 1.4 <sup>b</sup>
t <sub>lag</sub> (h)	$6.8 \pm 2.9^{a}$	$1.9 \pm 0.6^{b}$
t½ <sub>B</sub> (d)	$15.9 \pm 2.3^{a}$	$16.5 \pm 0.4^{a}$
MRT (d)	$13.3 \pm 0.83^{a}$	$13.2 \pm 0.09^{a}$

 $<sup>^{</sup>a,b}$ Values with different superscripts are significantly different ( $P \leq 0.05$ ).

the benzimidazoles, although increased plasma concentrations occur due to the increased rate of absorption from the abomasum, the bioavailability as measured by AUC is generally lower due to absence of the reservoir effect of the rumen resulting in a large reduction in residence time of these agents in the body<sup>14,25</sup>. The long elimination half-life of the halogenated salicylanilides, in contrast, is associated with their extensive plasma binding<sup>22</sup>, and therefore the residence time of rafoxanide in the body is not dependent on the reservoir effect of the rumen. The bioavailability of ivermectin, on the other hand, is significantly reduced after i.r. treatment and is attributed to intraruminal degradation of the agent<sup>27</sup>. Biodegradation of rafoxanide in the rumen has not been reported, although it has been shown to be more than twice as effective against F. hepatica when administered i-a as compared to either oral or i.r. treatment. This increased

efficacy of rafoxanide against *F. hepatica* is most likely due to an increase in drug plasma concentrations after i-a compared to i.r. administration and not due to differences in AUC. According to Maes *et al.*<sup>19</sup>, the flukicidal effect of the halogenated salicylanilide, closantel, is related more to peak plasma concentrations and less to residual persistent effect.

The study clearly showed the effect of the rumen on the rate of gastrointestinal absorption of rafoxanide. Further studies are required to examine the effect of type of feed, reduction in feed intake and transient feed withdrawal on the pharmacokinetics of the halogenated salicylanilides in sheep and to examine the influence of the rumino-reticulum on the disposition of these agents.

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#### **REFERENCES**

- Anon. 1989 SAS/STAT User's Guide. SAS Institute Incorporation, Cary, North Carolina
- Austin F H 1967 Absorption distribution, and excretion of sulphonamides in ruminants. Federation Proceedings 26: 1001–1005
- Barth D 1974 Rafoxanid Ein neues Leberegelmittel bei Schaf und Rind. Tierärztliche Umschau 29: 104–110
- Blood D C, Radostits O M 1989 Veterinary medicine. Bailliére Tindall, The University Press, Oxford
- 5. Bogan J, Marriner S 1987 The rumen as a pharmacokinetic compartment. In: Ooms L, Degryse A, Van Miert A S J P A M (eds) *Physiological and pharmacological aspects of the reticulo-rumen*. M A Kluver, Hingham: 253–269
- Boray J C, Roseby F B 1969 The effect of route of administration on the efficiency of clioxanide against immature Fasciola hepatica in sheep. Australian Veterinary Journal 45: 363–365
- Campbell N J, Brotowidjoyo M D 1975 The efficiency of clioxanide and rafoxanide against Fasciola hepatica in sheep by different routes of administration. Australian

- Veterinary Journal 51: 500-503
- Church D C 1988 The ruminant animal: digestive physiology and nutrition. Prentice Hall, New Jersey
- 9. Dedek W, Schwarz H, Liebaug E 1976 Untersuchungen zum Abbau und zur Ausscheidung des Anthelminthikums 131J-Rafoxanid an Rind und Schaf. Archiv für Experimentelle Veterinärmedizin 30: 423–426
- Dobson A 1967 Physiological peculiarities of the ruminant relevant to drug distribution. Symposium on Comparative Pharmacology. Federation Proceedings 26: 994–1000
- 11. Gibaldi M, Perrier D 1992 *Pharmacokinetics* (2nd edn). Marcel Dekker, New York
- 12. Guerrero J 1984 Closantel: a review of its antiparasitic activity. *Preventive Veterinary Medicine* 2: 317–327
- 13. Hall C A, Kelly J D, Whitlock H V, Ritchie L 1981 Prolonged anthelmintic effect of closantel and disophenol against a thiabendazole selected resistant strain of *Haemonchus contortus* in sheep. *Research in Veterinary Science* 31: 104–106
- 14. Hennessy D R 1994 The disposition of antiparasitic drugs in relation to the development of resistance by parasites of livestock. *Acta Tropica* 56: 125–141
- 15. Hennessy D R, Sangster N C, Steel J W, Collins G H 1993 Comparative pharmacokinetic disposition of closantel in sheep and goats. *Journal of Veterinary Pharmacology and Therapeutics* 16: 254–260
- Hinchcliff K W, Jernigan A D, Upson D W, Constable P D 1991 Ruminant pharmacology. Veterinary Clinic of North America: Food Animal Practice 7: 633–649
- 17. Jenkins W L, Davis L E, Boulos B M 1975 Transfer of drugs across the ruminal wall in goats. *American Journal of Veterinary Research* 36: 1771–1776
- 18. Lee RM 1973 The relationship between biological action and structure of some fascioli-

- cidal salicylanilides. Parasitology 67: 13–14
- Maes L, Lauwers H, Deckers W, Vanparijs O 1988 Flukicidal action of closantel against immature and mature Fasciola hepatica in experimentally infected rats and sheep. Research in Veterinary Science 44: 229–232
- Marriner S, Bogan J A 1979 The influence of the rumen on the absorption of drugs: studies using meclofenamic acid administered by various routes to sheep and cattle. *Journal of Veterinary Pharmacology and Therapeutics* 2: 109–115
- 21. Michiels M, Meuldermans W, Heykans J 1987 The metabolism and fate of closantel (Flukiver) in sheep and cattle. *Drug Metabolism Reviews* 18: 235–251
- 22. Mohammed-Ali N A K, Bogan J A 1987 The pharmacodynamics of the flukicidal salicylanilides, rafoxanide, closantel and oxyclosanide. *Journal of Veterinary Pharmacology and Therapeutics* 10: 127–133
- Nelder J A, Mead R 1965 A simplex method for function minimization. Computing Journal 7: 308–313
- Prichard R K 1978 Anthelmintics. In Proceedings, Refresher course for veterinarians Refresher course in the therapeutic jungle. University of Sydney: 39: 421–463
- 25. Prichard R K, Hennessy D R, Steel J W, Lacey W 1985 Metabolite concentrations in plasma following treatment of cattle with five anthelmintics. *Research in Veterinary Science* 39: 173–178
- Prichard R K, Steel J W, Hennessy D R 1981
  Fenbendazole and thiabendazole in cattle:
  partition of gastrointestinal absorption and
  pharmacokinetic behaviour. Journal of Veterinary Pharmacology and Therapeutics 4:
  295–304
- 27. Prichard R K, Steel J W, Lacey E, Hennessy D R 1985 Pharmacokinetics of ivermectin in sheep following intravenous, intraabomasal or intraruminal administration.

- Journal of Veterinary Pharmacology and Therapeutics 8: 88–94
- 28. Prins R A 1987 Rumen microbial metabolism of plant secondary compounds, xenobiotics and drugs. In: Ooms L, Degryse A, Van Miert A S J P A M (eds) *Physiological and pharmacological aspects of the reticulo-rumen*. M A Kluver, Hingham: 199–225
- 29. Rosenberger G 1973 Clinical examination of cattle. Paul Parey, Berlin.
- 30. Swan G E 1998 The pharmacology of halogenated salicylanilides and their anthelmintic use in animals. *Journal of the South African Veterinary Association* 70: 61–70.
- 31. Swan G E, Botha C J, Taylor J H, Mülders M S G, Minnaar P P, Kloeck A 1995 Differences in the oral bioavailability of three rafoxanide formulations in sheep. *Journal of the South African Veterinary Association* 66: 197–201
- 32. Swan G E, Mülders M S G 1993 Pharmacokinetics of rafoxanide in suckling and weaned lambs following oral administration. *Journal of the South African Veterinary* Association 64: 67–70
- 33. Swan GE, Schröder J 1981 A safety trial with rafoxanide in sheep. *Journal of the South African Veterinary Association* 52: 123–125
- 34. Symonds L E A, Roseby F B 1969 The use of clioxanide as an anthelmintic in sheep infected with *Haemonchus contortus*. *Australian Veterinary Journal* 45: 385–385
- 35. Taylor S M, Mallon T R, Blanchflower W J, Kennedy D G, Hewitt S A 1993 Effects of dietary variations on plasma concentrations of oral flukicides in sheep. *Journal of Veterinary Pharmacology and Therapeutics* 16: 48–54
- 36. Yamaoka K, Nakagawa T, Uno T 1978 Statistical moments in pharmacokinetics. *Journal of Pharmacokinetics and Biopharmaceutics* 6: 547–558