Prevalence of microorganisms associated with udder infections in dairy goats on small-scale farms in Kenya

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ABSTRACT

Six hundred and thirty clinically-normal milk samples from dairy goat flocks comprising a mixed population of German Alpine, Toggenburg, Saanen and Galla crosses were examined over a 3-month period to determine the prevalence of bacterial organisms. Bacteria were isolated in 28.7 % of the milk samples (181/630) either singly (92.8 %) or in combination (7.2 %). The most prevalent bacterial organisms were *Staphylococcus* spp. (60.3 %), followed by *Micrococcus* spp. (17.7 %), *Acinetobacter* spp. (5 %), *Actinomyces* spp. (5%) and *Streptococcus* spp. (1.1 %). The *Staphylococcus* spp. were mainly coagulase negative (64.3 %). Coagulase-negative staphylococci and coagulase-positive staphylococci accounted for 37.5 % and 22.7 % respectively of the total bacteria isolated. The isolation of bacteria, some of which are important in clinical and subclinical mastitis, in apparently normal caprine milk, indicates that particular attention should be given to the management of these dairy goat flocks in order to avoid the development of cases of clinical mastitis.

Key words: dairy goats, Kenya, microorganisms, udder infection.

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This study was carried out to determine the prevalence of intra-mammary bacterial infections in healthy dairy goats in the central Kenyan highlands, where the highest density of dairy goats in Kenya occurs, in order to formulate mastitis control measures in the flocks.

The investigation was carried out in 7 dairy goat farmer groups. All the does that were in different stages of lactation from all the dairy goat groups were sampled once a month for 3 months. The number of does sampled were 130 in the 1st month, 108 in the 2nd month and 77 in the 3rd month. The decrease in the number of does sampled in the 2nd and 3rd months was due to drying off of some of the does. The does were mainly German Alpine crosses with a few Toggenberg, Saanen and Galla crosses. The mammary glands of the does were examined visually for any injuries or swellings and by palpation for consistency and warmth. The first streams of milk from each mammary gland half were macroscopically examined. After discarding the first 3 streams of the fore milk, a 20 ml milk sample was collected aseptically from each mammary gland half into sterile Bijoux bottles. The milk samples were kept at 4 °C until bacteriologically processed in the laboratory between 3 and 6 h after collection. In the laboratory the milk samples were streaked onto sheep blood and MacConkey agar plates and incubated aerobically at 37 °C for 48 h. Significant bacterial colonies at 24 and 48 h were selected and sub-cultured for 24 h after which they were Gramstained and biochemically tested and classified according to standard methods⁵. With a few exceptions, most organisms were identified to genus level. The data were analysed using Statistix (SX v. 4.0) (Analytical Software, St Paul).

Visual evaluation and palpation of the mammary gland halves and the macroscopic examination of the milk revealed no abnormalities. The prevalence of bacteria in the 630 clinically normal milk samples cultured was 28.7 % (Table 1). The prevalence of bacteria isolated from normal caprine milk varies among studies undertaken in different countries³. This has been attributed to the influence of factors such as differences in the time of sampling, the diagnostic criteria used, breed differences, different hygiene and management practices followed on each farm, age and parity of the animals, and

the milking method^{2-4,11}. The goats used in this study were mainly stall-fed and were washed with warm water before being hand-milked. The does were milked twice a day, with no teat dipping after milking. Hand-milked goats have been found to have a lower prevalence of bacteria than machine-milked goats¹⁰. Hand milking of the does in this study may therefore have influenced the prevalence of bacteria. There were significantly (P < 0.05) more bacterial organisms isolated from the left mammary gland halves (55.8 %) than from the right mammary gland halves (44.2 %). This was similar to observations by Boscos et al.2, who suggested that this was possibly related to the milking process. They observed that most milkers were righted-handed and consequently milked the right mammary gland halves more efficiently that the left halves, hence predisposing the latter to a higher rate of bacterial infection.

Staphylococcus spp. were the most prevalent micro-organisms, accounting for 60.3 % (109/181) of all the bacterial isolates. Within this group, the coagulasenegative staphylococci were the most prevalent and comprised 62.4 (68/109) of the staphylococcal organisms isolated and 37.6 % (68/181) of the total bacterial isolates. This was followed by coagulasepositive staphylococci, which accounted for 37.5 % (41/109) of the staphylococci and 22.7 % (41/181) of the total bacterial isolates. Staphylococci are the most frequently-isolated microorganism from milk samples from both infected and non-infected glands¹⁻³, Staphylococcus aureus being the most commonly-isolated bacterium in clinically-infected caprine milk samples¹⁻³ The high incidence of coagulase-negative staphylococci observed was similar to other observations on non-clinical intra-mammary infections of goats 1-3,7,8. This bacterial organism, although not a major pathogen of clinical mastitis in goats, has been shown to persist throughout the lactation and dry periods, irritating the gland and causing a decrease in production and even clinical

The second most common group of microorganisms was *Micrococcus* spp.,

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Table 1: The prevalence of microorganisms isolated from normal caprine milk samples in Kenyan dairy goats.

Organism	1st sampling (<i>n</i> = 130) ^a		2nd sampling (<i>n</i> = 108) ^a		3rd sampling (<i>n</i> = 77) ^a		All samplings (<i>n</i> = 630) ^b
	Right	Left	Right	Left	Right	Left	Both halves
Nil	95 (73) ^e	82 (62)	81 (75)	76 (70)	59 (77)	56 (73)	449 (71.3) ^f
CNS ^c	15 (12)	16 (12)	12 (11)	9 (8)	9 (12)	7 (9)	68 (37.6) ^g
CPS ^d	6 (5)	7 (5)	7 (7)	9 (8)	5 (6)	7 (9)	41 (22.7) ^g
Micrococcus spp.	8 (6)	13 (10)	1 (1)	4 (4)	2 (3)	4 (5)	32 (17.7) ^g
Acinetobacter spp.	2 (2)	1 (1)	2 (2)	2 (2)	0 (0)	2 (3)	9 (5.0) ^g
Actinomyces spp.	0 (0)	3 (4)	1 (1)	3 (3)	2 (3)	0 (0)	9 (5.0) ^g
Streptococcus spp.	1 (1)	0 (0)	0 (0)	0 (0)	0 (0)	1 (1)	2 (1.1) ^g
Other spp.	1 (1)	3 (4)	1 (1)	2 (2)	0 (0)	0 (0)	7 (3.9) ^g
CNS + Micrococcus spp.	1 (1)	1 (1)	1 (1)	2 (2)	0 (0)	0 (0)	5 (2.7) ^g
Actinomyces + others	1 (1)	1 (1)	1 (1)	0 (0)	0 (0)	0 (0)	3 (1.6) ^g
Acinetobacter + others	0 (0)	2 (2)	1 (1)	1 (1)	0 (0)	0 (0)	4 (2.2) ^g
CPS + others	0 (0)	1 (1)	0 (0)	0 (0)	0 (0)	0 (0)	1 (0.5) ^g

a,bNumber of quarter milk samples (a) for either (right or left) mammary gland half and (b) for all mammary gland halves.

which accounted for 17.7 % of all bacterial isolates. The other bacterial organisms isolated were Acinetobacter spp. 5 %, Actinomyces spp. 5 %, Streptococcus spp 1.1 % and mixed bacteria (7.2 %). Actinomyces spp. and Streptococcus spp. are important causative organisms of bovine mastitis^{6,9}. The absence of infected cattle in the environment of the does may thus have contributed to the low prevalence of these organisms in the dairy goats examined. The high prevalence (28.7 %) of bacterial isolates from apparently normal milk samples indicates a high percentage of subclinical infections in these dairy goat flocks. It also demonstrates that presence of bacterial organisms in the mammary gland halves of dairy goats does not necessarily result in clinical mastitis. However, the isolation of microorganisms from apparently normal caprine milk, some of which are important in clinical and subclinical mastitis (especially Staphylococcus spp.), indicates that particular attention should be given to the management of these dairy goat flocks to avoid cases of clinical mastitis. This also emphasises the need for

adequate pasteurisation of the milk before consumption.

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CNS = coagulase-negative Staphylococcus.

^dCPS = coagulase-positive *Staphylococcus*.

e.f.gNumber in brackets is percentage (e) of the number of the quarter milk samples, (f) total quarter milk samples (630), and (g) of the total bacterial isolate (181).