# Renal involvement in dogs with babesiosis

R G Lobetti<sup>a\*</sup> and L S Jacobson<sup>a</sup>

### **ABSTRACT**

Proteinuria, and renal tubular casts and epithelial cells in urine sediment, are commonly observed in both complicated and uncomplicated babesiosis, but do not necessarily reflect or predict renal failure. This study investigated the presence and degree of renal damage in canine babesiosis. Renal function and integrity were evaluated using serum urea and creatinine, serum electrolytes (sodium and potassium), fractional clearance of sodium  $(Fc_{Na})$  and potassium  $(Fc_K)$ , urine enzyme activity of gamma-glutamyl transpeptidase and alkaline phosphatase, urine protein:creatinine ratio, and urinalysis. One control group (n = 10) and 3 groups of babesiosis cases were studied: mild uncomplicated (n = 10), severe uncomplicated (n = 11), and complicated (n = 9). All babesiosis groups showed wellconcentrated urine. Mean serum urea was elevated in the severe and complicated groups, and was significantly different from the control group. There was no statistically significant difference between the groups for creatinine, although the complicated group had a mean value above the normal reference range. Hypokalaemia was uncommon in all the groups. Hyperkalaemia was present in only 2 dogs in the complicated group. Marginal hyponatraemia was present in a minority of dogs in all groups. The serum electrolytes were not significantly different between groups. There was no overall elevation, nor any statistically significant difference in both the FcNa and FcK between the groups. Only 1 dog, in the complicated group, showed marked enzymuria. Proteinuria was a common finding and was significantly different between the severe and complicated groups and the control group. Some dogs in all groups had renal tubular epithelial cells in the urinary sediment, which increased in severity from the mild to the complicated groups and was significantly different from the control group. This study demonstrated that minimal renal damage occurs more often in canine babesiosis than significant damage or acute renal failure.

Key words: babesiosis, canine, kidney, renal function.

Lobetti R G, Jacobson L S Renal involvement in dogs with babesiosis. *Journal of the South African Veterinary Association* (2001) 72(1): 23–28 (En.). Department of Companion Animal Medicine, Faculty of Veterinary Science, University of Pretoria, Private Bag X04, Onderstepoort, 0110 South Africa.

## INTRODUCTION

Canine babesiosis is an important worldwide tick-borne disease caused by the intra-erythrocytic protozoal parasites Babesia canis or B. gibsoni<sup>39</sup>. Although the disease primarily involves erythrocyte destruction, it may also result in multisystemic involvement<sup>39,40</sup>. Acute renal failure (ARF) is an uncommon complication of babesiosis and typically presents as anuria or oliguria despite adequate rehydration<sup>21</sup>. Evidence of renal damage, reflected on urinalysis by the presence of proteinuria, casts and renal tubular epithelial cells, is common in both complicated and uncomplicated cases, but does not necessarily reflect or predict renal failure<sup>26</sup>. In babesiosis, elevated serum

Received: September 2000. Accepted: February 2001.

urea alone is an unreliable indicator of renal insufficiency, as a disproportionate rise in urea, compared with creatinine, occurs, possibly due to catabolism of lysed erythrocytes<sup>33</sup>. Renal failure is diagnosed on the basis of ongoing evaluation of urine volume, urinalysis and degree of azotaemia. In humans, falciparum malaria, a disease clinically similar to canine babesiosis<sup>25</sup>, can result in ARF, which resembles sepsis-related acute tubular necrosis<sup>6</sup>. Glomerulonephritis may also be evident<sup>6</sup>.

Primary or intrinsic ARF is a syndrome characterised by the sudden onset of impaired renal function, resulting in azotaemia, fractional clearance of sodium (Fc<sub>Na</sub>) that is greater than 1, the presence of renal tubular epithelial (RTE) cells and/or casts in the urine sediment, and characteristic histological changes<sup>1,4,11,13</sup>. Any toxic or ischaemic renal insult may result in cellular degeneration and/or necrosis, with consequent RTE cell loss

into the urine. In humans, overt necrosis is not evident in all cases but tubular dysfunction is a uniform hallmark of this form of ARF<sup>1</sup>. Ischaemic injury occurs when renal blood flow is attenuated by decreased blood pressure or renal vasoconstriction<sup>44</sup>. Glomerular afferent arteriolar vasoconstriction caused by the effects of angiotensin II and antidiuretic hormone (ADH), in response to increased renin release, is a proposed mechanism of decreased glomerular filtration rate (GFR) in ARF<sup>28</sup>. Decreased renal blood flow results in reduced amounts of oxygen and metabolic substrates presented to tubular cells, and this 'cellular starvation' initiates the development of acute tubular necrosis with consequent ARF<sup>5</sup>. Acute renal failure associated with malaria has been attributed to hypovolaemia and/or hypotension, intravascular haemolysis, hyperparasitaemia, cholestatic jaundice, catecholamines, and endotoxemia<sup>31,36</sup>.

In canine babesiosis, the morphological lesions in the kidney have been attributed to anaemic hypoxia resulting from erythrocyte destruction<sup>17</sup>. However, recent unpublished data, from 84 dogs with complicated babesiosis, have shown that the mean haematocrit of dogs with elevated creatinine was significantly higher (mean 36.5 %, SD 20.19) than of those with normal creatinine (mean 22 %, SD 16.38), making hypovolaemia a more likely cause than anaemia for the renal failure described in this disease. Babesiosis can result in a kidney that is swollen and dark in colour, with redbrown urine in the bladder. Microscopically the RTE cells are swollen and contain haemoglobin (Hb) droplets and small vacuoles. In severe cases, necrosis of the RTE cells is evident. The lumen of the nephron contains multiple Hb casts<sup>17</sup>. The net effect of babesiosis on the kidney can be ARF, which has been attributed to haemoglobinuric nephropathy<sup>26</sup>. However, ARF is uncommon in babesiosis 17,39 and recent work has demonstrated that severe haemoglobinuria, of the magnitude seen in canine babesiosis, did not induce significant nephropathy, regardless of whether or not concomitant anaemia was present<sup>24</sup>

The true pathogenesis of renal lesions in

failure<sup>26</sup>. In babesiosis, elevated serum

\*\*Department of Companion Animal Medicine, Faculty of Veterinary Science, University of Pretoria, Private Bag X04, 0110 Onderstepoort, South Africa.

<sup>\*</sup>Author for correspondence. Bryanston Veterinary Hospital, PO Box 67092, Bryanston, 2021 South Africa. E-mail: rlobetti@mweb.co.za

babesiosis is still obscure. Maegraith<sup>25</sup> noted the development of oliguria or anuria in dogs without concomitant haemoglobinuria. Several authors have reported that anoxia, reduction in renal blood flow, and possibly hypotension with intra-renal vasoconstriction and renal ischaemia must be considered of major importance in the pathogenesis, as opposed to mechanical obstruction of tubules by Hb and the toxic effects of Hb<sup>25,26,46</sup>. It has also been demonstrated that hypoxia results in more injury to renal tubules than haemoglobinuria and that the nephrotoxic effect of Hb appears to be highly individual<sup>24</sup>. Malherbe<sup>26</sup> also suggested that the renal damage in babesiosis is usually reversible.

The purpose of this study was to investigate the presence and degree of renal damage in naturally occurring canine babesiosis. Renal function and integrity were evaluated using urinalysis, serum urea and creatinine, serum electrolytes (sodium and potassium), fractional clearance of sodium (Fc<sub>Na</sub>) and potassium (Fc<sub>K</sub>), urine enzyme activity of gammaglutamyl transpeptidase (GGT) and alkaline phosphatase (ALP), and quantification of proteinuria.

### **MATERIALS AND METHODS**

# Study design

The Ethics and Research Committees of the Faculty of Veterinary Science, University of Pretoria, approved this study and written consent by the dogs' owners was obtained. Thirty dogs with babesiosis, presented to the Onderstepoort Veterinary Academic Hospital (OVAH), were sequentially enrolled. The diagnosis of babesiosis was based on finding B. canis parasites on a thin capillary blood smear, stained with Cams Quick stain (C A Milsch). These dogs were categorised into 3 groups: mild uncomplicated (Group 1). severe uncomplicated (Group 2) and complicated (Group 3). Mild cases had mild-to-moderate anaemia (haematocrit 20-30 %) with no clinical or biochemical signs of complicated disease. Severe cases had severe anaemia (haematocrit < 15 %) with no clinical or biochemical signs of complicated disease. Complicated cases had one or more of the following complications: cerebral signs, ARF, acute respiratory distress syndrome, hypotensive shock, or haemoconcentration. Clinically healthy, aparasitaemic, dogs, presented to the OVAH for routine ovariohysterectomy, were used as controls (Group 4). Groups 1 and 4 comprised 10 dogs each and groups 2 and 3, 11 and 9 dogs respectively.

### Data collection

Blood was collected from the cephalic vein, using a 22G venoject needle, a holder and a serum vacuum tube (Vacutainer System, Becton Dickinson). A cystocentesis urine sample was collected aseptically using a 23G needle and a 10-m  $\!\ell$ syringe. All samples were collected before any treatment.

### Analytical methods

Urea and creatinine were determined on a Technicon RA 1000 system (Technicon Instruments Corporation) using the Technicon modification of the kinetic method for urea<sup>42</sup> and the alkaline picrate reaction for creatinine, modified as a first order rate reaction<sup>19</sup> Urine and serum sodium and potassium were determined using an ion selective analyser (Nova 1, Nova Biomedical). Urine ALP and GGT were determined on a Technicon RA 1000 system using the Technicon modification of the p-nitrophenyl phosphate substrate method in AMP buffer for ALP41 and glutamyl-p-nitroanilide substrate with glycylglycine peptide acceptor for GGT<sup>38</sup>. Urine Hb was determined on a Technicon RA 1000 system using the Drabkins method (C A Milsch). Total urine protein was determined using a spectrophotometer (Lange LP6 Photometer) based on the Richterich technique using perchloric acid and biuret reagent<sup>18</sup>. Urine Hb was subtracted from the total urine protein and the corrected protein value was expressed as a ratio with the urine

The fractional clearance of sodium and potassium was calculated, using the following formula:

 $\frac{\text{Urine electrolyte}}{\text{Serum electrolyte}} \times \frac{\text{Serum creatinine}}{\text{Urine creatinine}}$ 

The physicochemical evaluation of the urine was performed using a urine dipstick (Lenstrip 8 Dipsticks, Benmore Diagnostics) and an AO veterinary refractometer (American Optical, Scientific Instrument division). Microscopic evaluation was done on urine sediment stained with Sternheimer-Malbin stain (Kyron Laboratories), which enabled the differentiation of RTE cells from other urinary epithelial cells<sup>3</sup>. The presence of RTE cells in the urine was subjectively scored on a scale of 1-4 as follows: 1, represented 1 RTE cell per 2-3 high power fields (HPF); 2, represented 1-2 RTE cells per HPF; 3, represented 2-4 RTE cells per HPF, and 4, represented more than 5 RTE cells per HPF.

# Data analysis

Statistical analysis of the data was performed using a commercial statistical

software package (Sigma Stat, Jandel Scientific Software). Parameters that were statistically analysed were urine specific gravity (SG), serum urea and creatinine, serum sodium and potassium, Fc<sub>Na</sub>, Fc<sub>K</sub>, urine ALP and GGT activity (expressed as a ratio to urine creatinine), proteinuria (expressed as urine protein: creatinine ratio), and urinalysis findings (haemoglobinuria and RTE cells in sediment). Data were compared between the groups using analysis of variance (ANOVA). The urinalysis findings were compared using Friedman repeated measures ANOVA on ranks. The Tukey correction was used for group comparisons. The Pearson test was used to check for correlation between haemoglobinuria and serum creatinine concentrations and urine RTE cell score. In all analyses, a value of P < 0.05 was considered significant.

### **RESULTS**

Clinicopathological findings are summarised in Table 1.

Most dogs in the severe and complicated groups (8/11 and 8/9 respectively) had elevated serum urea, compared with only 1/10 in the mild group (Fig. 1). The serum creatinine did not mirror the serum urea in that 1/10 in the mild group, 0/10 in the severe group, and 3/10 in the complicated group had elevated serum creatinine levels (Fig. 2). There was a significant positive correlation between serum urea and creatinine in the mild (r =0.89, P < 0.05) and complicated (r = 0.83, P< 0.05) groups, but not in the severe group.

One dog in each of the babesiosis groups showed mild hypokalaemia. Severe hypokalaemia was present in 1 dog in the severe and 1 in the complicated group. Hyperkalaemia was present in 1 dog in the complicated group. There was no statistically significant difference between any of the groups for either hypo- or hyperkalaemia. Marginal hyponatraemia was present in 2/10 in the mild and severe groups, and 3/10 in the complicated group. There was no statistically significant difference between any of the groups. None of the dogs showed hypernatraemia. There was no elevation in the Fc<sub>Na</sub> indicative of acute tubular dysfunction, but there was a statistically significant difference between all 3 groups and the control group in that the babesiosis groups had a much lower Fc<sub>Na</sub> than the control group. There was neither elevation of, nor any statistically significant differences for, Fc<sub>k</sub>.

The urine ALP: creatinine ratio was elevated in 1 dog in the mild group and 1 in the severe group, and 2 in the complicated group. The urine GGT: creatinine

Table 1: Clinicopathological parameters in dogs with mild, severe, and complicated babesiosis and normal control dogs.

Parameter	Mild (n = 10)		Severe ( <i>n</i> = 11)		Complicated $(n = 9)$		Control $(n = 10)$	
	Median	Range	Median	Range	Median	Range	Median	Range
Serum urea (mmol/t)	5.9	3.3–15.3	12.8	7.3–29	19.3	5.5–25.1	6.8	3.7–12.6
Serum creatinine (µmol/ℓ)	98	74–168	93	38–130	125	59–281	119.5	105–155
Serum sodium(mmol/ℓ)	141.5	138–145	142	139–150	142	133–149	17.5	141–152
Serum potassium (mmol/ℓ)	3.85	3.6-4.0	3.9	2.8-5.0	4.3	2.4-6.6	4.4	4.2-4.7
Urine ALP: creatinine ratio	0.27	0.2-3.41	0.51	0.16-2.42	0.14	0.07-46.91	0.5	0.25-1.17
Urine GGT: creatinine ratio	0.57	0.28-2.05	1.27	0.24-4.03	1.11	0.22-6.7	0.86	0.48-1.78
Fc <sub>Na</sub>	0.17	0-0.67	0.06	0-0.27	0.05	0-0.22	0.49	0-1.4
Fc <sub>K</sub>	11.69	4.33-29.88	20.42	2.88-43.88	28.09	8.98-43.05	16.26	7.84-27.76
Urine haemoglobin (g/t)	1.25	0.2-2.5	1.6	0.3-11.6	1.95	0.92-3.80	0.65	0.3-3.4
Urine protein: creatinine ratio	0.08	0.01-0.98	0.77	0.04-2.66	1.24	0.08-3.78	0.08	0.01-0.40
Specific gravity	1.05	1.04-1.06	1.05	1.03-1.06	1.05	1.035-1.06	1.043	1.03-1.06
RTE cells	0.5	0–1	1	0–2	2	0–4	0	0–0

ratio was elevated in 1 dog in the mild group, 2/11 in the severe group, and 2/9 in the complicated group. There was, however, no correlation between the ALP and GGT creatinine ratios nor was there any statistical difference between the groups for urine ALP and GGT activity (Fig. 3). Only 1 dog in the complicated group showed marked enzymuria (both ALP and GGT), but this was not associated with azotaemia.

An elevated urine protein: creatinine ratio was evident in 3/10 of the mild group, 6/11 of the severe group, and 8/9 of the complicated group. When compared with the control group, the proteinuria was statistically significant in the severe and complicated groups (Fig. 4). There was no difference in the urine SG between the groups. All dogs had well-concentrated urine (>1.030). Haemoglobinuria was present in all babesiosis

groups with medians of 1.25, 1.65 and 3.95 g/l for the mild, severe and complicated groups, respectively. There was no correlation between the haemoglobinuria and serum creatinine concentrations or urine RTE cell score. Some dogs in all the babesiosis groups had RTE cells in the sediment, which increased in severity from the mild to the complicated groups (Fig. 5). Tubular casts (cellular or granular) were not evident in any of the dogs.

Four dogs showed a number of clinicopathological changes consistent with acute renal damage (1 in the mild and 3 in the complicated groups). The dog in the mild group had moderate azotaemia, mild elevation in the urine enzymes, and moderate proteinuria. Hyperkalaemia, elevated  $F_{C_{Na}}$ , or RTE celluria were not present. In the complicated group, the 3 dogs had moderate azotaemia and moderate RTE celluria. One of these had

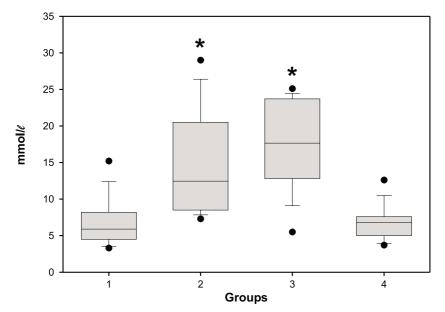


Fig. 1: Serum urea for the 4 groups. Data are shown as median (horizontal line within box), 25th and 75th percentiles (horizontal ends of boxes), and 10th and 90th percentiles (T-bars). An asterisk indicates significant differences between the babesiosis groups and the control group. Black dots represent outliers.

hyperkalaemia and hyponatraemia, and 2 had moderate proteinuria and enzymuria. None of the dogs showed elevated  $Fc_{Na}$ . All 4 dogs responded well to standard therapy for  $ARF^{13}$ .

### **DISCUSSION**

In this study, the most consistent finding indicative of renal damage was proteinuria, and its severity was related to the severity of the babesiosis. Renal and renal-related disorders due to human falciparum malaria, a disease similar to canine babesiosis<sup>25</sup>, include both extrarenal and renal manifestations<sup>6</sup>. The former consist of fluid and electrolyte disorders. The latter vary widely, ranging from mild proteinuria and glomerulonephritis to ARF<sup>8,36</sup>. In humans the reported incidence of ARF associated with falciparum malaria ranges from 1–4 %8. Jacobson and Clark<sup>21</sup> reported that ARF is an uncommon complication in canine babesiosis, being diagnosed in only 3 of 134 cases reviewed, giving an incidence of 2.2 %.

The elevated serum urea in the severe uncomplicated group in this study, without a corresponding increase in the serum creatinine, confirms previous findings<sup>33</sup>. The phenomenon has been attributed to catabolism of lysed erythrocytes, resulting in an increased ammonia load on the liver and consequent increased urea production<sup>33</sup>, but could also be associated with more generalised protein catabolism resulting from a febrile, inflammatory illness. Other possible causes would include gastrointestinal haemorrhage and ingestion of a high protein meal. These latter causes are unlikely in canine babesiosis. Thus elevated serum urea alone is an unreliable indicator of renal insufficiency in babesiosis. Elevated serum urea in the complicated group in this study was associated with a concomitant increase in the serum creatinine,

which could reflect decreased renal blood flow, possibly as a result of decreased blood pressure and/or hypovolaemia. A recent study showed that hypotension occurred frequently in canine babesiosis and the presence and severity of hypotension increased with increased disease severity<sup>22</sup>. As elevated creatinine, evident in some of the cases, was not correlated with any of the other parameters of renal function or integrity, it may have been pre-renal in origin. Another study has shown that elevated creatinine is associated with increased risk of death in canine babesiosis<sup>45</sup>, indicating that it might nonetheless be a useful measure of renal insufficiency. In babesiosis, a higher cut-off value and/or serial creatinine determinations would assist in distinguishing prerenal from renal causes.

Urine enzyme activity is both an early and persistent indicator of renal tubular damage<sup>34</sup>. Both GGT and ALP are brush border enzymes present in the proximal convoluted tubule of the kidney<sup>14</sup>. Although 24-hour urine enzyme activity is more accurate, evaluation of the ratio of urine enzyme: creatinine in a spot urine sample is technically simpler and has been shown to correlate well with a 24-hour sample<sup>34</sup>. Urine ALP and GGT activity  $> 10 \text{ U}/\ell$  and a ratio >2 (calculated using SI units) can be considered to be elevated 12,16,34. Only 1 dog in the complicated group revealed severe changes in the urine enzyme activity; however, this was not accompanied by abnormal urine SG or Fc<sub>Na</sub>.

The Fc<sub>Na</sub> can be used as an indicator of acute tubular dysfunction, with an increase in the Fc<sub>Na</sub> over 1 reportedly indicating acute tubular dysfunction<sup>10</sup>. Two of the control dogs had  $Fc_{Na}$  greater than 1, raising the possibility that a Fc<sub>Na</sub> greater than 1 is not always indicative of acute tubular dysfunction. In this study an unexpected finding was that the mean Fc<sub>Na</sub> in the dogs with babesiosis was lower than that of the control dogs. Most sodium is actively re-absorbed from the proximal convoluted tubules of the kidneys, resulting in passive water reabsorption. Further sodium reabsorption occurs in the distal convoluted tubules (secondary to the active re-absorption of chloride ions) and collecting ducts (controlled by aldosterone)34. In this study, the lower FC<sub>Na</sub> can be interpreted as either renal retention of sodium secondary to aldosterone secretion or inhibition of prostaglandins, or as a result of activation of the renin-angiotensin-aldosterone system, in response to renal arterial hypotension<sup>15,27,30</sup>. The well-concentrated urine in all dogs with babesiosis can also be attributed to sodium and water

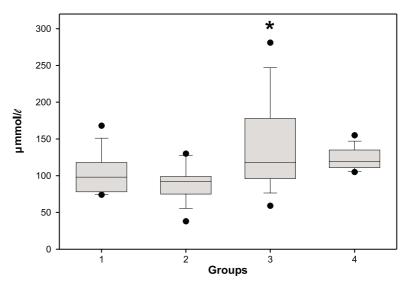


Fig. 2: Serum creatinine for the 4 groups. Data are shown as median (horizontal line within box), 25th and 75th percentiles (horizontal ends of boxes), and 10th and 90th percentiles (T-bars). Black dots represent outliers.

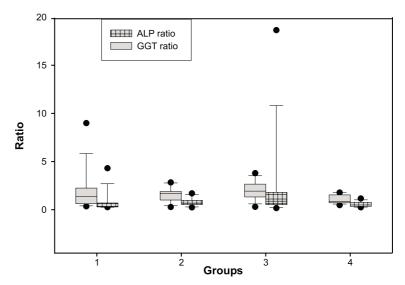


Fig. 3: Urine ALP: creatinine and GGT: creatinine ratio for the 4 groups. Data are shown as median (horizontal line within box), 25th and 75th percentiles (horizontal ends of boxes), and 10th and 90th percentiles (T-bars). Black dots represent outliers.

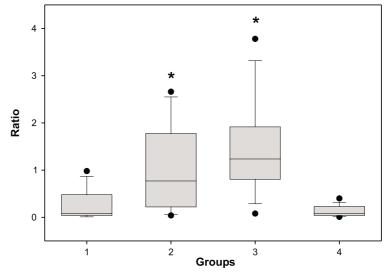


Fig. 4: Urine protein:creatinine ratio for the 4 groups. Data are shown as median (horizontal line within box), 25th and 75th percentiles (horizontal ends of boxes), and 10th and 90th percentiles (T-bars). An asterisk indicates significant differences between the babesiosis groups and the control group. Black dots represent outliers.

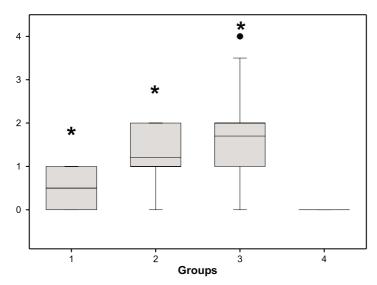


Fig. 5: Renal tubular epithelial cells in urine sediment for the 4 groups. Data are shown as median (horizontal line within box), 25th and 75th percentiles (horizontal ends of boxes), and 10th and 90th percentiles (T-bars). An asterisk indicates significant differences between the babesiosis groups and the control group. Black dots represent outliers.

retention. However, sodium retention may have been inadequate, as some of the dogs were hyponatraemic, which could be indicative of early renal tubular dysfunction.

Hyponatraemia, usually asymptomatic, can occur in up to 67 % of patients with malaria without renal failure and is often associated with severe disease<sup>36</sup>. In children, hyponatraemia occurred during the acute phase of the disease with increased fractional sodium excretion and continuing sodium wastage in 17 % after recovery<sup>9,37</sup>. In the current study, hyponatraemia was present in only 17 % of the cases and Fc<sub>Na</sub> was not increased in any of the dogs. In patients with falciparum malaria it has been shown that there is a decreased response to water load attributed to peripheral vasodilatation, which results in a decreased effective blood volume leading to the release of vasopressin and norepinephrine, increased renin activity and decreased renal perfusion<sup>36</sup>. It has also been suggested that water retention occurs in babesiosis<sup>35</sup>. Hyponatraemia in malaria has been ascribed to multiple factors: increased secretion of ADH, hypervolaemia, and accumulation of sodium in both parasitised and non-parasitised red blood cells as a result of decreased sodium-potassium ATP-ase activity<sup>36</sup>.

Hypokalaemia is an uncommon finding in malaria, as it was in this study. In malaria, hypokalaemia has been ascribed to decreased potassium intake and respiratory alkalosis<sup>6</sup>, both of which can occur in babesiosis<sup>23</sup>. The 5 dogs that revealed hypokalaemia had a normal to increased Fc<sub>k</sub>, which is an abnormal response and could thus be indicative of early renal tubular dysfunction or as a response to

the alkalosis.

Hyperkalaemia occurs in malaria patients with either ARF or intravascular haemolysis<sup>36</sup>. In this study, 2 dogs showed hyperkalaemia, of which 1 had ARF. In a study in children, plasma potassium was significantly higher and the Fc<sub>k</sub> significantly lower during the acute illness than after recovery<sup>37</sup>. As canine red blood cells are much lower in potassium than human red blood cells, hyperkalaemia is unlikely to occur as a result of haemolysis in the dog.

In this study, a number of dogs had RTE cells in the urine sediment. In malaria, abnormal urinary sediment, consisting of red and white blood cells and occasional granular casts, commonly occurs in patients without renal failure<sup>36</sup>. Other urinary sediment abnormalities, such as the presence of RTE cells, are not commonly reported<sup>29</sup>. The presence of RTE cells in the urine sediment can indicate renal damage due to hypoxia, hypoperfusion, or toxic damage. In this study, the number of RTE cells in the urine sediment increased with increased disease severity. Renal hypoxia results in rounding and retraction of RTE cells with a disruption of actin microfilaments, as a result of which a large number of viable RTE cells are sloughed into the urine<sup>32</sup>. Acute renal hypoxia has also been shown to induce apoptotic changes<sup>20</sup>. This may explain the lack of correlation between urine enzyme activity and the presence of RTE cells.

Proteinuria was a common finding in this study. Proteinuria was demonstrated by Maegraith<sup>25</sup> in experimental babesiosis, and occurred within 24 hours of the first appearance of parasites in the blood. In later stages of the infection, there was evidence of tubular damage in the form of granular and hyaline casts. Cast formation was not evident in this study. Mild proteinuria occurred in 40 % of children with malaria during the acute illness but was not related to creatinine clearance, body temperature at presentation, or peripheral parasite density. Proteinuria was also absent after recovery<sup>37</sup>. Babesia rodhaini-infected mice developed immune-complex-induced mesangiopathic glomerulonephropathy and moderate renal tubular necrosis2. These mice had elevated serum urea and proteinuria. Babesia microti-infected mice showed relatively mild immunecomplex-induced mesangiopathic glomerulonephropathy and mild renal tubular necrosis, with no increase in serum urea and no proteinuria<sup>2</sup>. The degree of proteinuria in this study was consistent with tubulo-interstitial disease<sup>7,10</sup>. Canine babesiosis has not been associated with glomerulonephropathy<sup>7</sup>.

Acute renal failure in malaria is commonly associated with blackwater fever, characterised by fever, massive intravascular haemolysis and haemoglobinuria<sup>6,43</sup>. The condition is associated with quinine administration in patients with glucose-6-phosphate dehydrogenase (G6PD) deficiency<sup>36,43</sup>. The presence or absence of G6PD deficiency in dogs with babesiosis has never been evaluated, but severe haemoglobinuria is unlikely to mimic the human situation, since the common antibabesial drugs are dissimilar to quinine and haemoglobinuria generally occurs before treatment. In our experience, dogs with babesiosis, that die as a result of ARF show severe haemoglobinuria and oliquria, and frequently produce a small volume of urine that is almost black in colour. In this study there was no correlation between the haemoglobinuria and serum creatinine and RTE cells in the urine. This supports a previous study that showed that haemoglobinuria does not induce renal failure<sup>24</sup>.

### CONCLUSION

This study demonstrated RTE celluria, variable enzymuria, proteinuria, and variable azotaemia in dogs with babesiosis. However, these were all minimal changes and all could be consistent with hypoxia and/or hypovolaemia. To fully elucidate the effect of babesiosis on the kidney, further studies involving histological assessment are needed.

### **ACKNOWLEDGEMENTS**

This study was partially funded by the Pet Memorial fund of the South African Veterinary Foundation. The authors would like to thank the laboratory technicians of the Clinical Pathology Laboratory, Department of Companion Animal Medicine, Faculty of Veterinary Science, University of Pretoria, for their valuable assistance. Dr Jacobson was supported by a grant from the Technology and Human Resources for Industry Programme (THRIP) of the Department of Trade and Industry and National Research Foundation.

### **REFERENCES**

- Anderson R J, Schrier R W 1991 Acute RE-NAL Failure. In Wilson J D, Braunwald E, Isselbacher K J, Petersdorf R G, Martin J B, Fauci A S, Root R K (eds) Harrison's principles of internal medicine (12th edn). McGraw-Hill, New York: 1144–1150
- 2. Annable C R, Ward P A 1974 Immunopathology of the renal complications of babesiosis. *Journal of Immunology* 112: 1–8
- Anon. 1961 Atlas of Use of Sternheimer-Malbin Staining Technique. Abbot Laboratories, Chicago, USA
- Behrend E N, Grauer G F, Mani I 1996. Hospital-acquired acute renal failure in dogs: 29 cases (1983–1992). Journal of the American Veterinary Medical Association 208: 537–541
- Brown S A 1993 Physiology of the urinary tract. In Slatter D (ed.) Textbook of small animal surgery. W B Saunders, Philadelphia: 1384–1395
- Day N P, Phu N H, Loc P P 1997 Malaria and acute renal failure. Journal of the Royal College of Physicians of London 31: 146–148
- DiBartola S P, Chew D J, Jacobs G 1980 Quantitative urinalysis including 24-hour protein excretion in the dog. *Journal of the American Animal Hospital Association* 16: 537–546
- 8. Eiam-Ong S, Sitprija V 1998 Falciparum malaria and the kidney: a model of inflammation. *American Journal of Kidney Diseases* 32: 361–375
- English M C, Waruiru C, Lightowler C 1996 Hyponatraemia and dehydration in severe malaria. Archives of Diseases of Children 74: 201–205
- Finco D R 1995 Urinary protein loss. In Osborne C A, Finco D A (eds) Canine and feline nephrology and urology. Lea and Febiger, Baltimore: 211–215
- Finn W E 1990 Diagnosis and management of acute tubular necrosis. Medical Clinics of North America 74: 873–879
- 12. Gossett K A, Turnwald G H, Kearney M T 1987 Evaluation of γ-glutamyl transpeptidase-to-creatinine ratio from spot samples of urine supernatant, as an indicator of urine enzyme excretion. *American Journal of Veterinary Research* 48: 455–457
- 13. Grauer G F, Lane I F 1995 Acute renal failure: Ischemic and chemical nephrosis. In Osborne C A, Finco D R (eds) *canine and feline nephrology and urology.* Lea and Febiger, Baltimore: 441–459

- Guder W G, Ross B D 1984 Enzyme distribution along the nephron. Kidney International 26: 101–111.
- 15. Guyton A C 1991 Textbook of medical physiology (6th edn). W B Saunders, Philadelphia: 420–434.
- Heiene R, Biewenga W J, Koeman J P 1991 Urinary alkaline phosphatase and gamma glutamyl transpeptidase as indicators of acute renal damage in dogs. *Journal of Small Animal Practice* 32: 521–524
- Hildebrandt P K 1981 The organ and vascular pathology of babesiosis. In Ristic M, Koeier I P (eds) Babesiosis. Academic Press, New York: 459–473
- 18. Hoffmann J.P. Richterich R 1970 Elimination of turbidity during determination of plasma proteins with the biuret reagent. Zeitschrift für Klinisie Chemie und Klinisie Biochemie 8: 595–598
- 19. Jaffe M Z 1986 Ueber den Niederschlag. Welchen pikrinsaure in normalen erzeugt und ueber eine neue Reaction des Kreatinins. Zeitschnft für Physiologische Chemie 10: 391–400
- 20. Jaffe R, Ariel I, Beeri R 1997 Frequent apoptosis in human kidneys after acute renal hypoperfusion. *Experimental Nephrology* 5: 399–403
- 21. Jacobson L J, Clark I 1994 The pathophysiology of canine babesiosis: new approaches to an old puzzle. *Journal of the South African Veterinary Association* 65: 134–145
- Jacobson L S, Lobetti R G, Vaughan-Scott T 2000 Blood pressure changes in dogs with babesiosis. Journal of the South African Veterinary Association 71: 14–20
- Leisewitz A L, Jacobson L S, Reyers F 1999 Mixed acid-base disturbances of severe canine babesiosis. In Proceedings of a Symposium on Canine Babesiosis and Ehrlichiosis, Onderstepoort, Pretoria, 6 November 1999: 37–44
- 24. Lobetti R G, Reyers F, Nesbit J W 1996 The comparative role of haemoglobinuria and hypoxia in the development of canine babesial nephropathy. *Journal of the South African Veterinary Association* 67: 188–198
- 25. Maegraith B, Gilles H M, Devakul K 1957 Pathological processes in *Babesia canis* infections. *Zeitschrift für Tropenmedizen und Parasitologie* 8: 485–514
- 26. Malherbe W 1966 Clinico-pathological studies of *Babesia canis* infection in dogs. The influence of the infection on kidney function. *Journal of the South African Veterinary Association* 37: 261–264
- Mason D E 1993 Anesthesia and the urinary system. In Slatter D (ed.) Textbook of small animal surgery. W B Saunders, Philadelphia: 2267–2271
- Monroe W E, Waldron D R. 1993 Renal failure: surgical considerations. In Bojrab M I (ed.) Disease mechanisms in small animal surgery (2nd edn). W B Saunders, Philadelphia: 417–425.
- 29. Obatomi D K, IJkole C O 1996 Increased excretion of urinary enzymes in patients

- with renal disorders. *Medical Science Research* 24: 529–531
- Power I, Cumming A D, Pugh G C 1992 Effects of diclofenac on renal function and prostacylin generation after surgery. British Journal of Anaesthesiology 69: 5: 451–456
- 31. Prakash J, Gupta A, Kumar O 1996 Acute renal failure in falciparum malaria; increasing prevalence in some areas of India; a need for awareness. *Nephrology, Dialysis and Transplantation* 11: 2414–2416
- 32. Racusen L C 1998 Epithelial cell shedding in acute renal injury. *Clinical and Experimental Pharmacology and Physiology* 25: 273–275
- 33. Reyers F 1992 Is the azotaemia in canine babesiosis an indication of renal disease. Proceedings of the 9th Faculty Day, University of Pretoria, Faculty of Veterinary Science, 1 October 1992: 17
- 34. Rivers B J, Walter P A, O'Brien T D 1996 Evaluation of urine gamma-glutamyl transpeptidase-to-creatinine ratio as a diagnostic tool in an experimental model of aminoglycoside-induced acute renal failure in the dog. Journal of the American Hospital Association 32: 323–336
- Schetters T P M, Kleuskens J, Scholtes N 1998 Parasite localization and dissemination in the Babesia-infected host. Annals of Tropical Medicine and Parasitology 92: 513–519
- 36. Sitprija V 1988 Nephropathy in falciparum malaria. *Kidney International* 34: 867–877
- Sowunmi A 1996 Renal function in acute falciparum malaria. Archives of Diseases of Children 74: 293–298
- 38. Szasz G 1976 Reaction rate method for gamma glutamyltransferase activity in serum. *Clinical Chemistry* 22: 2051–2055
- Taboada J, Merchant J 1991. Babesiosis of companion animals and man. Veterinary Clinics of North America: Small Animal Practice 21: 103–123
- Taboada J 1998 Babesiosis. In Greene C G (ed.) Infectious diseases of the dog and cat (2nd edn). W B Saunders, Philadelphia: 473–481
- 41. Tietz W 1980 Progress in the development of a recommended method for alkaline phosphatase activity measurements. *Clinical Chemistry* 26: 1023–1035
- 42. Tiffany T O, Jansen J M, Burtis C A 1972 Enzymatic kinetic rate and end-point analyses of substrate, by use of a GeMSAEC fast analyzer. *Clinical Chemistry* 18: 829–840
- Trang T H, Day N P, Ly V C 1996 Blackwater fever in southern Vietnam: a prospective descriptive study of 50 cases. Clinical Infectious Diseases 23: 1274–1281
- Wilkes B M, Mialloux L U 1986 Acute renal failure; pathogenesis and prevention. American Journal of Medicine 80: 1129–1136
- 45. Van Zyl M 1995 Prediction of survival in hospitalised cases of canine babesiosis. MMedVet thesis, University of Pretoria
- Zwart D, Brocklesby D W 1979 Babesiosis: non-specific resistance, immunological factors and pathogenesis. Advances in Parasitology 7: 49–113