



A comparison of mycotoxin contamination of premium and grocery brands of pelleted cat food in South Africa

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Contamination with mycotoxins is of concern to pet owners and veterinary practitioners owing to their ability to cause disease and exacerbate the pathological changes associated with other diseases. Currently, there is a lack of information regarding the mycotoxin content of common premium brand (PB) and grocery brand (GB) cat feeds. Therefore, we undertook to determine the mycobiota content of feed samples, from both categories ($n = 6$ each), and measured the levels of aflatoxin (AF), fumonisin (FB), ochratoxin A (OTA) and zearalenone (ZEA) by high performance liquid chromatographic analysis. There were high concentrations of mycotoxins in both categories of feed, regardless of the notion that PBs are of a higher quality. The concentration of these toxins may contribute to the development of related pathologies in felines.

Introduction

Mycotoxins have been implicated in adverse effects in both human and animal health (Fink-Gremmels 1999; Pulina et al. 2014). In a worldwide survey (2004–2011) of over 17 000 samples of feed or feed ingredients, it was found that more than 75% of samples were contaminated by at least one mycotoxin and 40% of the samples contained at least two mycotoxins (Streit et al. 2013). Currently, about 300 mycotoxins have been identified but not all are necessarily implicated in toxicity. The Food and Agriculture Organization (FAO) estimates that a quarter of the food produced globally is contaminated with mycotoxins. This causes significant economic losses as well as poses a serious threat to human and animal health (Bryden 2012; Vasanthi & Bhat 1998). Hence, regulatory limits have been recommended by organisations such as the Food and Drug Administration (FDA) for the common mycotoxins. Mycotoxins commonly implicated in and associated with animal health concerns include aflatoxin (AF), fumonisin (FB), ochratoxin A (OTA), trichothecenes and zearalenone (ZEA) (Boermans & Leung 2007).

Dry, pelleted pet food often contains 5% – 25% of animal protein or its derivatives with the remaining ingredients consisting of corn, corn gluten, wheat, wheat gluten and rice and its by-products, amongst other 'millings' (Klich & Pitt 1988). In a highly competitive pet food market, cost-cutting exercises are inevitable, leading to a compromise in the quality of products entering the retail sector. These cereal products that are often unfit for human consumption can act as excellent substrates for fungal proliferation and production of mycotoxins that contribute to liver, kidney and other diseases in pets (Bucci et al. 1998; Dereszynski et al. 2008). It is the contamination of cereals at harvest, post-harvest, manufacture and then storage (Bennett & Klich 2003; Tulpule 1981) that often becomes a health risk to pets by causing mycotoxicosis incidents and death. In 2011, South Africa experienced an outbreak of aflatoxicosis as a result of the consumption of poor quality, low-cost pelleted food (Arnot et al. 2012). The exacerbating factor was mouldy and low-grade peanuts that were contaminated with *Aspergillus flavus* and *Aspergillus parasiticus*.

In this study, we compared the mycotoxin profiles of premium brand (PB) and grocery brand (GB) cat food. PB products are perceived to have low amounts of cereal whilst GBs are perceived to have higher cereal content. Though no major mycotoxin outbreaks have been recorded in felines in recent years, the implication of mycotoxins and their role in feline health cannot be ignored (De Souza & Scussel 2012). Examination of cat food labelling on packaging reveals that claims of high crude protein contents refer largely to vegetable and animal by-products and minimally to meat. Packaging labels provide extensive information on ingredients but limited information on actual percentages of ingredients in the formulation, a trend that is seen in both market segments and often leads to misunderstanding amongst consumers with regard to the nutritional value of the product. Furthermore, information gained from this study may warrant further investigation and contribute to consumer knowledge and feline health.

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Materials and methods

Materials

Chemicals, reagents and mycotoxin standards were obtained from Merck (South Africa) and Sigma (South Africa) unless otherwise specified. All mycotoxin standards, except the fumonisins, were purchased from Sigma (St. Louis, USA), whilst fumonisin B₁ (FB₁) and FB₂ were purchased from PROMEC (MRC, South Africa). For this study, PB refers to all veterinary restricted brands that may be purchased at veterinary practices or retail veterinary shops (Vetshops) only and generally are priced between R80.00 and R120.00 per kilogram, whilst GBs are commonly sold in supermarket and grocery outlets at a lower price range between R30 and R60 per kilogram.

Methodology

Sampling

Pelleted cat food ($n = 12$) from two marketing channels (PB and GB) were selected for this study. Samples were purchased from their respective outlets in convenient sizes of 2 kg – 3 kg packets. Information on brand, package size, expiry date and barcode serial numbers were recorded. Each packet of food was emptied into a 5-L bucket and thoroughly mixed by shaking. The sampling technique was adapted from methods described by Tittlemier et al. (2011). The bucket was divided into quadrants and approximately 125 g per quadrant sample was scooped up with a clean metal ladle. The samples were thoroughly mixed prior to obtaining a representative sub-sample of 500 g of which a further sub-sample of 200 g was taken by dividing 500 g into four sub-samples and 50 g taken from each quadrant. All feed samples (200 g each) were milled to a fine powder using a mechanical blender (Petron 3600, Germany). The milled samples were used for fungal culture and mycotoxin determination. Remaining samples were resealed and stored in sealed containers at 4 °C until required for further analysis.

Fungal isolation: Fungal isolation as well as subculturing and subsequent identification of fungi were done as previously described (Kaufman, Williams & Sumner 1963; Singh & Chuturgoon 2017).

Mycotoxin extraction and clean-up of feed samples: Mycotoxin extractions were done as described (Singh & Chuturgoon 2017). Mean recoveries are provided in Table 1.

Thin layer chromatography: Thin layer chromatography (TLC) was run for each mycotoxin as previously described (Singh & Chuturgoon 2017).

TABLE 1: Mean recoveries of selected mycotoxins after spiking in feed samples using high performance liquid chromatography.

| Mycotoxin | Concentration spiked (µg/kg) | Concentration measured (µg/kg) | % of recovery |
|------------------|------------------------------|--------------------------------|---------------|
| AFB ₁ | 100 | 95.5 | 95.5 |
| AFB ₂ | 100 | 89.0 | 89.0 |
| OTA | 100 | 94.6 | 94.6 |
| ZEA | 100 | 93.0 | 93.0 |
| FB ₁ | 200 | 196.4 | 98.2 |
| FB ₂ | 200 | 193.0 | 96.5 |

High performance liquid chromatographic analysis of feed sample extracts: High performance liquid chromatographic (HPLC) analysis of feed sample extracts was performed as previously described (Singh & Chuturgoon 2017).

Results

Thin layer chromatography characterisation and HPLC quantitation (µg/mL) were performed for the commonly suspected mycotoxins implicated in pet food contamination, namely, AF, FB, OTA and ZEA (Liggett et al. 1986; Shephard & Sewram 2004; Stenske et al. 2006). The most prevalent fungal isolates in all samples were *Aspergillus* species, *Fusarium* species and *Penicillium* species (Table 2). These fungi were found in both PB and GB feed categories. The fungal species *Aspergillus flavus*, *Aspergillus fumigatus* and *Aspergillus niger* were more commonly isolated while *A. parasiticus*, *Aspergillus ochraceus*, *Aspergillus poae* and *Aspergillus penicillioides* were found less commonly in the samples tested.

Using TLC, all samples in both categories tested positive for four mycotoxins (Table 3). The PB samples appeared to fare worse than GB samples, particularly in terms of AF and ZEA concentrations. HPLC analysis investigated AF for AFB₁ and AFB₂, while FB was evaluated for FB₁ and FB₂ besides OTA and ZEA. Both PB and GB failed the limits set by the *Fertilizer, Farm Feeds, Agricultural Remedies and Stock remedies Act* (No. 36 of 1947) of 10 ppb (1 ppb = 1 µg/L) for total AFs (South African Government 2009). The levels of AFs (Table 4)

TABLE 2: Fungal species identification and selected mycotoxin detection in premium brand and grocery brand cat pelleted feed samples.

| Fungal species | Fungal sub-species | Fungal isolates (CFU's/mL) | |
|--------------------|---------------------------|----------------------------|---------|
| | | Premium | Grocery |
| <i>Aspergillus</i> | <i>A. flavus</i> | *** | ** |
| | <i>A. fumigatus</i> | * | ** |
| | <i>A. niger</i> | ** | ** |
| | <i>A. niveus</i> | - | - |
| | <i>A. ochraceus</i> | - | ** |
| | <i>A. parasiticus</i> | * | ** |
| | <i>A. penicillioides</i> | * | * |
| | <i>A. poae</i> | ** | - |
| <i>Fusarium</i> | <i>F. graminearum</i> | ** | *** |
| | <i>F. verticillioides</i> | * | * |
| <i>Penicillium</i> | <i>Penicillium</i> spp. | ** | * |
| | <i>P. polonicum</i> | - | * |
| | <i>P. crustosum</i> | - | * |
| Other | <i>Rhizopus</i> spp. | + | - |
| | Unidentified microbe | + | - |
| | Yeast | + | + |

A., *Aspergillus*; F., *Fusarium*; P., *Penicillium*.

*, 100–300 × 10⁴ CFUs; **, 300–500 × 10⁴ CFUs; ***, > 500 × 10⁴ CFUs; +, positive only.

TABLE 3: The results of thin layer chromatography characterisation.

| Mycotoxin | TLC characterisation | |
|--------------|----------------------|---------|
| | Premium | Grocery |
| Aflatoxin | *** | ** |
| Fumonisin | * | ** |
| Ochratoxin A | ** | ** |
| Zearalenone | ** | * |

TCL, thin layer chromatography.

***, very intense spot; **, intense spot; *, spot; (intensity of spot as compared to a standard of each mycotoxin).

TABLE 4: The results of the high performance liquid chromatographic quantitation of mycotoxins in cat feed extracts.

| Feed market segment | AFB ₁ | AFB ₂ | FB ₁ | FB ₂ | OTA | ZEA |
|---------------------|------------------|------------------|-----------------|-----------------|------|------|
| Premium brand | 125.02** | 11.77* | 9.98 | 5.45 | 1.32 | 9.1* |
| Grocery brand | 41.57 | 6.30 | 202.53*** | 118.37*** | 0.74 | 2.27 |

Note: For statistical significance, data are expressed as mean \pm s.d.

OTA, ochratoxin A; ZEA, zearalenone.

*, $p < 0.05$; **, $p < 0.005$; ***, $p < 0.0001$

detected in the PB were over the set limits for both AFB₁ (125.02 ppb) and AFB₂ (11.77 ppb) but GB only exceeded the limits for AFB₁ (41.57 ppb). The amounts of both AFB₁ ($p = 0.0087$) and AFB₂ ($p = 0.0091$) in PB were statistically significantly higher as compared to GB. *Fusarium graminearum* was predominantly isolated in both categories; however, HPLC analysis indicated that the GB had exceedingly high concentrations of both FB₁ (202.53 ppb) and FB₂ (118.37 ppb), failing the limit set by the Food and Drug Administration of 100 ppb (FDA 2001). The amounts of both FB₁ ($p = 0.028$) and FB₂ ($p = 0.0041$) were significantly higher in GB as compared to PB. OTA ($p = 0.0196$) and ZEA ($p = 0.0060$) levels were significantly higher in PB as compared to GB (Table 4).

In summary, the PB fared worse than the GB in its AF, OTA and ZEA contamination, whilst GB contained much higher levels of FB than PB.

Discussion

Cats are obligatory carnivores and require taurine in their diets. A good animal protein source will provide the taurine required for a cat's good health. The presence of high amounts of mycotoxins in commercial cat diets is indicative of high cereal content. PBs are perceived as better quality with superior nutrition than GBs, but they present a mycotoxin risk due to their high levels of cereal content. A study in Brazil showed a high correlation between diets rich in grains with a reduced immunity to infections in domestic animals (De Souza & Scussel 2012). Clinical signs described for dogs with aflatoxicosis are depression, anorexia, weakness, icterus and sudden death (Arnot et al. 2012; Ketterer et al. 1975; Stenske et al. 2006). Cats with sub-acute aflatoxicosis often show signs of lethargy, anorexia and progressive weight loss. Cats have lower susceptibility to mycotoxicosis than dogs but continuous exposure to low concentrations of mycotoxins in the feed can induce accumulative effects, leading to chronic liver and kidney damage (Dereszynski et al. 2008; Patterson & Roberts 1979).

The PB samples revealed a higher count of *A. flavus* colony forming units (CFUs) than the GB samples. This finding is, however, not surprising as these are ubiquitous soil fungi and common contaminants of corn, groundnuts and other cereal grains used in animal feed production (Leung, Díaz-Llano & Smith 2006) and often implicated in aflatoxicosis. In addition, *Fusarium graminearum* was detected at higher concentrations in the GB samples and are associated with FB, fusaric acid and ZEA production. *Penicillium* species were less commonly noted but PB samples showed higher

concentrations than the GB samples. *Penicillium* spp. produces tremorgenic mycotoxins such as roquefortine and penitrem A. These mycotoxins induce tremorgenic mycotoxicosis particularly in canines characterised by acute abdominal pain, salivation, vomiting, fever, muscle tremors with hyperaesthesia, seizures and sometimes even death (Hocking, Holds & Tobin 1988; Naudé et al. 2002; Young et al. 2003). OTA is produced by a number of *Aspergillus* and *Penicillium* species while ZEA is produced by *Fusarium* species (Leung et al. 2006; Shephard & Sewram 2004). OTA and ZEA were detected (by TLC and quantified by HPLC) at very low levels, but various studies have described toxicity at these reported levels (Leung et al. 2006). The role of *Fusarium* mycotoxins in animal health is particularly important economically as they have been implicated in infertility and reproductive dysfunction in sheep, cattle and pigs. Poultry are particularly affected with loss in weight, egg production and gastrointestinal lesions (Antonissen et al. 2014; D'Mello, Placinta & Macdonald 1999; Placinta, D'Mello & Macdonald 1999).

A mycotoxin mix of FB₁, FB₂, OTA and ZEA together with AFs may present a higher risk to illness or mycotoxicosis. Many researchers have reported the simultaneous occurrence of several mycotoxins in feed and feed ingredients (Fox, Hodgkins & Smart 2012; Mwanza 2007; Tulpule 1981). This potent mycotoxin combination may result in synergistic action and potentiate effects that support the multi-aetiological theory (Boermans & Leung 2007; Creppy et al. 2004; Mwanza et al. 2013; Ryu, Jackson & Bullerman 2002).

Irrespective of marketing channels, all products were contaminated with mycotoxins. The mean AF concentration across the various brands indicates that all products failed the prescribed limit (10 ppb; by the *Fertilizer, Farm Feeds, Agricultural Remedies and Stock remedies Act* [No. 36 of 1947]; South African Government 2009). The long-term exposure of cats to mycotoxins may be implicated in numerous clinical conditions such as neoplasia, reduced immunity and poor growth and fertility (De Souza & Scussel 2012).

Conclusion

PBs are marketed as superior feeds, but their cereal content makes them susceptible to mycotoxin contamination. Many PBs are imported and the higher mycotoxin content may be attributed to lengthy transport in containers on the high seas and high humidity. Though cats may appear to be less susceptible to mycotoxicosis, the risk of long-term exposure to mycotoxins coupled with poor health or concurrent disease could result in increased susceptibility. Further *in vivo* studies are required to evaluate feline susceptibility to mycotoxins.

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Competing interests

The authors declare that they have no financial or personal relationships that may have inappropriately influenced them in writing this article.

Authors' contributions

S.D.S. performed experiments, analysed data and prepared draft manuscript. S.B. assisted with analyses and preparation of the manuscript. A.A.C. was the supervisor of S.D.S. for a PhD and assisted with data analysis and drafting of the final manuscript.

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