

Changes in Mithun (*Bos frontalis*) spermatozoa during epididymal passage

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ABSTRACT

Genital organs of 10 healthy, adult Mithun bulls (6–8 years old) that were slaughtered at the dwellings of tribal people for meat were collected. Immediately after collection, spermatozoa from 3 different regions of the epididymis, i.e. the head, body and tail, were obtained to study morphological changes of the spermatozoa during passage through these regions. The prevalence of proximal cytoplasmic droplets significantly decreased from the head to the tail of the epididymis. Conversely, the percentage of distal cytoplasmic droplets increased significantly from the head to the tail region. The incidence of tailless heads rose significantly from head to body and then reduced significantly in the tail region. The percentage of total head abnormalities did, however, not change markedly, but total mid-piece and tail abnormalities differed significantly between the three epididymal regions.

Keywords: epididymal passage, male reproduction, Mithun, sperm morphology.

Bhattacharyya H K, Bujarbaruah K M, Goswami B K, Biswas R K, Khan I **Changes in Mithun (*Bos frontalis*) spermatozoa during epididymal passage.** *Journal of the South African Veterinary Association* (2010) 81(2): 80–81 (En.). Teaching Veterinary Clinical Complex, Faculty of Veterinary Sciences and Animal Husbandry, Shuhama-Alusteng, Srinagar – 190 006, Jammu and Kashmir, India.

epididymal fluid containing spermatozoa from these regions was suspended separately in buffered formal saline. The suspended fluid was further suitably diluted in buffered formal saline to facilitate the study of sperm characteristics, namely proximal cytoplasmic droplets, distal cytoplasmic droplets, tailless heads and mid-piece and tail abnormalities, using a phase contrast microscope. Sperm head abnormalities were studied under oil immersion ($\times 100$) using a light microscope. The epididymal fluid suspension was used to prepare the thin smears stained with Carbol-fuchsin-eosin stain^{17,11}. Hundreds of spermatozoa per region of each epididymis were examined. The Mann-Whitney test was used to test for differences between the right and left parts of the epididymis, while the Kruskal-Wallis test was used to test for differences between the head, body and tail of the epididymis.

RESULTS

As the spermatozoa pass through the epididymis there is a successive and significant reduction in the prevalence of proximal cytoplasmic droplets between the head, body and tail, and droplets were virtually absent in the tail (Table 1). On the other hand, the prevalence of distal cytoplasmic droplets increased significantly in succession in the 3 epididymal regions. The occurrence of tailless heads rose significantly from head to body and then decreased significantly in the tail region (Table 1). Percentage of total sperm head abnormalities did not differ significantly between the 3 regions. Abnormalities of the mid-piece and tail were absent in the head and increased significantly from there to the body and further to the tail. No significant difference was found between the right and left parts of the epididymis.

DISCUSSION

The increase in the prevalence of distal cytoplasmic droplets from the head to the tail regions of the epididymis corresponds to the migration process of the cytoplasmic droplet from a proximal to distal position during epididymal transit. The significant decrease in proximal cytoplasmic droplet and significant increase in distal cytoplasmic droplet percentages

INTRODUCTION

The Mithun (*Bos frontalis*) is a semi-wild bovid¹³. The World Conservation Monitoring Cell lists this 'ceremonial ox' of north-eastern mountainous regions of India as endangered. It plays an important role in the socioeconomic life of the tribal communities but is yet to be scientifically studied in detail⁴. The Mithun is usually kept as a free-ranging animal in forests. Tribal people slaughter the Mithun for its meat, which they consider to be precious.

The number of Mithuns has declined greatly in recent times, creating the need to identify fertile males to use as sires in order to conserve the species. In the bull, the most striking morphological change in the spermatozoa during their passage through epididymis is the migration of the cytoplasmic droplet from the proximal to the distal end of the mid-piece^{9,12}. Other changes have also been described^{3,8}. Knowing the morphological changes in spermatozoa during epididymal passage

in the Mithun will help define semen quality in the species. Thus the present study examines morphological changes as the spermatozoa pass through the epididymis.

MATERIALS AND METHODS

The present study was conducted in the National Research Centre on Mithun (NRCM), Indian Council of Agricultural Research (ICAR), Nagaland, India. Genital organs of 10 healthy, adult Mithun bulls aged 6–8 years were collected from the dwellings of tribal people when they sacrificed them during marriage ceremonies. The animals were found to be in good health and no physical abnormalities were noticed pre- or post-slaughter. Different parts of the external and internal genitalia were also free of gross pathological changes.

The study was conducted from January to mid June. Maximum and minimum temperatures varied between 21.0 and 9.4 °C in January and between 30.0 and 25.71 °C in June. Monthly rainfall was 9.4–289.5 mm and relative humidity 72.5–83.8%. Immediately after collection the genital organs were transported to the laboratory in normal saline solution at 37 °C. The epididymis was dissected from the testis and epididymal fluid was obtained by incision with a scalpel followed by gentle pressure on the central part of each of three regions of the epididymis, namely the head, body and tail. The

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Received: September 2008. Accepted: June 2009.

Table 1: Occurrence of sperm abnormalities (mean $\pm$ SE) in different regions of the epididymis in Mithuns (*Bos frontalis*).

	Head (%)	Body (%)	Tail (%)	P-value
Proximal cytoplasmic droplet	37.36 $\pm$ 2.22*	10.55 $\pm$ 0.61*	0.09 $\pm$ 0.09*	0.000
Distal cytoplasmic droplet	19.18 $\pm$ 0.92*	23.36 $\pm$ 1.91*	37.09 $\pm$ 0.09*	0.000
Tailless head	2.45 $\pm$ 0.67*	5.20 $\pm$ 1.60*	1.45 $\pm$ 0.40*	0.000
Total head abnormalities	1.45 $\pm$ 0.41	1.36 $\pm$ 0.41	1.67 $\pm$ 0.32	0.560
Total mid-piece abnormalities	0*	0.82 $\pm$ 0.33*	2.27 $\pm$ 0.45*	0.000
Total tail abnormalities	0*	0.09 $\pm$ 0.09*	1.45 $\pm$ 0.39*	0.001

*Significant at $P < 0.05$, SE = standard error.

in the spermatozoa from the head to the tail regions of the epididymis was also reported by earlier workers in bulls¹, rams², boars¹⁶ and goats¹⁵. The finding in the current study that the prevalence of tailless spermatozoa is higher in the body of the epididymis than in the head or tail is similar to that of Rao¹⁴, who showed a higher prevalence in the distal part of the head compared to the tail. Since the neck region of spermatozoa is vulnerable and fragile, the sperm head can become detached from the mid-piece with the initiation of motility, which takes place at the base of the sperm head⁵. Spermatozoa acquire motility between the head and tail of the epididymis and hence the scope for detachment of the head of the spermatozoa could be greatest in the lower head or body, which might explain the significantly higher incidence of tailless heads in the body region. The significant reduction in the incidence of tailless heads in the tail could be attributed to the dissolution and liquefaction³ or phagocytosis within the tail epididymis. In agreement with the current study showing no effect of epididymal passage on the prevalence of sperm head abnormalities, Igboeli¹⁰ observed that the prevalence of sperm head abnormalities is also similar in the head and tail of the epididymis of domestic bulls. The present finding in respect of tail abnormalities is in general agreement with observations of earlier workers^{2,6} that most of the tail abnormalities arise after the spermatozoa leave the testis.

Cury and Vinha⁷ also reported that the incidence of tail abnormalities increased with the passage of spermatozoa through the epididymis. The increase in the percentage of mid-piece and abnormalities in the tail compared to the body could be ascribed to the exposure of the spermatozoa to varying biochemical microenvironments during their passage through the epididymis that might influence the development of sperm tail abnormalities. The present study revealed that the changes in sperm characteristics during epididymal passage in Mithun bulls are similar to those previously recorded in bulls but with some variation.

ACKNOWLEDGEMENT

The authors are grateful to the Director, NRCM, ICAR, Jharnapani, Nagaland, for providing facilities.

REFERENCES

1. Amann R P, Almquist J O 1962 Reproductive capacity of dairy bulls. VII. Morphology of epididymal sperm. *Journal of Dairy Science* 45: 1516–1526
2. Amann R P, Hay S R, Hammerstedt R H 1982 Yield, characteristics, motility and cAMP content of sperm isolated from seven regions of ram epididymis. *Biology of Reproduction* 27: 723–733
3. Amann R P, Almquist J O 1962 Reproductive capacity of dairy bulls. VI. Effect of unilateral vasectomy and ejaculation frequency on sperm reserves; aspects of epididymal physiology. *Journal of Reproduction and Fertility* 3: 260–268
4. Bhattacharyya H K, Rafiqul Islam, Bujarbaruah K M 2005 Mithun: a unique large

5. ruminant of northeastern India. *Livestock International* 9: 22–23
5. Bhosrekar M R 1990 *Semen production and artificial insemination* (2nd edn). Privately published, Alahabad, India
6. Blom E 1950 Interpretation of spermatocytology in bulls. *Journal of Fertility and Sterility* 1: 223–238
7. Cury L T, Vinha N A 1982 Changes in the morphology of goat spermatozoa during their passage through the epididymis. *Arquivos da Escola de Veterinaria da Universidade Federal de Minas Gerais* 34: 267–271 (cf. *Animal Breeding Abstract* 51: 310)
8. Gustafson B 1966 Luminal contents of the bovine epididymis under conditions of reduced spermatogenesis, luminal blockage and certain sperm abnormalities. *Acta Veterinaria Scandinavica* 7: Suppl. 17
9. Hancock J I 1955 The disintegration of bull spermatozoa. *Veterinary Record* 67: 825–826
10. Igboeli G 1968 Bull and rabbit testes and epididymal spermatozoa at normal and body temperatures studied with the aid of ligatures, PhD thesis, Cornell University, Ithaca, New York
11. Lagerlof N 1934. Morphologische Untersuchungen über Veränderungen in spermabild und in den Hoden bei Bullen mit Verminderter oder aufgehobener Fertilität *Acta Pathology Microbiology Scandinavica* Suppl: 1–19
12. Nicander I 1958 Studies on the regional histology and cytochemistry of the ductus epididymis in stallions, rams and bulls. *Acta Morphology Nederland Scandinavia* 1: 337–362
13. Pathak N N, Barua K K 1980 The Mithun of Nagaland and Arunachal. *Science Report* 17: 483
14. Prasad C S, Reddy J M 2000 Morphological changes of goat spermatozoa during epididymal transit. *Indian Journal of Animal Reproduction* 21: 27–29
15. Rao A R 1971 Changes in the morphology of sperm during their passage through the genital tract in bulls with normal and impaired spermatogenesis. PhD thesis, Royal Veterinary College, Stockholm, Sweden
16. Wekerle L, Szollosi E, Sarlos P, Schnaider L 1984 Morphological examination of sperm cells obtained from the testes and epididymis of boars. *Magyar Allatorvosok Lapja* 40: 663–666 (cf. *Animal Breeding Abstract* 54: 53–56)
17. Williams W W 1920 Technique of collecting semen for laboratory examination with a review of several diseased bulls. *Cornell Veterinarian* 10: 87–94